

Brain aging research

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Introduction

The last three decades produced a striking increase in investigations of the neurobiological basis of brain aging and aging-related changes in neural and cognitive function. Experimental and clinical studies of aging have become more valuable as the population, at least in industrialized countries, has become 'greyer'. The increase in adult life expectancy that occurred in the twentieth century produced the motivation and necessity to invest resources in increasing 'health span' as well as lifespan, in order to maximize quality of life and minimize the financial and social burdens associated with disability in the later years of life. Specific interest in the aging nervous system is driven by recognition that increased longevity has little appeal for most people unless it is accompanied by maintenance of cognitive abilities. Indeed, surveys of older individuals routinely show that loss of mental capacity is among their greatest fear. In recent years, neuroscientists and gerontologists, with a variety of training and experimental approaches, have applied increasingly powerful quantitative methods to investigate why neural function declines with age. New animal model systems have been developed and old ones have become better characterized and standardized. The necessary and important descriptive studies that dominated the field in earlier years are increasingly supplemented by more hypothesis-driven research, resulting in sophisticated investigations and models of the mechanisms of brain aging. This review provides a selective overview of recent and current research on brain aging. The focus throughout will be on normal brain aging and the moderate cognitive changes that often accompany it, not on aging-related neurodegenerative diseases that result in

dementia. To provide a context for studies of neurobiological changes in the aging brain, a brief overview of the types of cognitive changes that are commonly seen in aging humans is first provided. The remainder of the review focuses on animal studies that are progressively overcoming the unique challenges of aging research to reveal the neurobiological mechanisms of aging-related cognitive dysfunction, and suggest new targets for therapies to prevent or ameliorate cognitive decline.

Cognitive changes in aging

The range of cognitive changes that occur with aging suggests that aging-related neurobiological changes are not limited to restricted neural regions, as occurs with some aging-related neurological diseases, but rather affect many and disparate areas of the brain. Nevertheless, variation among individuals in the types and extent of aging-related cognitive changes demonstrates that deficits in specific functions, mediated by different neural regions, can develop to some extent independently. The nature and range of aging-related cognitive deficits in humans has been well-described in a recent review and is discussed here only briefly.¹

Basic cognitive functions, such as attention and memory, can be significantly affected by aging but, in both cases, some aspects are typically maintained while others decline. Although there is variation across studies, selective attention and the ability to sustain attention appear to be well maintained across aging.^{2,3} Reports of aging-related deficits in visual-attention tasks and other measures of attention (such as the Stroop task) may be due to a general decline in the rate of information processing rather than specific deficits in selective attention.⁴ Although selective attention is maintained, many older individuals have deficits in tasks that require attention-switching or attending to and processing information from multiple sources of information.^{5,6} Such deficits

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appear to be reversible to some extent by training and even by exercise,⁷ although it is not clear whether such improvements involve reversal of neurobiological changes that led to the deficit or development of alternative strategies.

Deficits in memory are probably the aging-related change most commonly recognized by aging individuals and most widely investigated by experimental gerontologists.⁸ With respect to long-term memory,¹ aging significantly affects memory for specific events (episodic memory) whereas some other aspects of long-term memory, such as procedural memory, are well maintained. Clearly, many older individuals exhibit deficits in reorganizing and integrating information held in working memory.^{9–11} Such dysfunction may not necessarily reflect a problem with working memory per se, but rather may be one manifestation of aging-related problems in executive control. This range of neural processes is involved in allocating attention, inhibiting irrelevant information, directing problem-solving, and essentially planning and coordinating neural activities related to cognitive functions.^{2,12,13} These executive activities depend critically upon the proper function of the prefrontal cortex and its reciprocal connections with other cortical regions.¹⁴ The prominence of aging-related changes in executive function, coupled with prominent changes in frontal cortex that are evident in imaging and postmortem analyses of the human brain, are consistent with hypotheses that executive control deficits arising from changes in the frontal lobe explain much of cognitive aging.¹⁵ It is clear, however, that changes in other neural regions, such as the hippocampus, contribute as well (see below).

Investigating the mechanisms of aging-related cognitive decline

The careful elucidation of aging-related cognitive deficits in humans is critical for understanding the problems faced by older adults, but human studies provide only limited ability to investigate underlying neurobiological mechanisms. Until recently, investigations of neuroanatomical and neurochemical changes associated with human cognitive decline were limited to postmortem analyses. The final state of the brain could be assessed in detail but changes occurring as cognitive deficits developed could not. With

advances in brain imaging it now is possible to assess structural changes in individual brains as they age,^{16–18} and also to investigate aging-related changes in cerebral blood flow.^{19,20} Moreover, developments in magnetic resonance (MR) spectroscopy and positron emission tomography now permit analysis of metabolic changes and alterations in neurotransmitters and their receptors *in vivo*.^{21–26}

Despite such advances, experimental studies of the mechanisms of brain aging depend extensively on the use of animal models, particularly non-human primates and rodents. In the US, the importance of primate and rodent models has been recognized with extensive and critical financial support from the National Institute on Aging.²⁷ Clearly, one important consideration in the use of animal models is the extent to which they undergo aging-related cognitive changes that are similar to those that occur in humans. Rhesus monkeys have an extensive behavioral repertoire and can be trained in many tasks that assess memory and executive function in a manner analogous to human studies. Rhesus monkeys show very similar deficits in memory and executive function^{28,29} and, as in humans, the extent of cognitive dysfunction varies among individuals.^{30,31} Although it is more difficult to relate cognitive changes in humans to cognitive changes in rodents, learning and memory can be readily assessed using a variety of maze and other tasks.^{32–35} Similarly, methods now have been developed for testing executive function in mice and rats and demonstrate clear aging-related decline in such frontal cortex-dependent tasks.^{36–38} The demonstration that aging-related cognitive dysfunction is similar in humans and rodents has made it possible to claim the many experimental advantages provided by the latter, including the use of powerful molecular genetic techniques in mice.^{39–41} Thus, it is to studies in monkeys and rodents that we owe much of the current understanding of the mechanisms of normal brain aging.

Structural changes in the aging brain

Even a cursory comparison of histological sections or brain images from a young adult versus an aged individual makes it clear that the structure of the human brain changes with age, even in the absence of neurodegenerative disease. Typically, the aged

brain exhibits regionally-specific reduction in the volume of both grey and white matter.^{42,43} Consistent with the view that functions mediated by the frontal lobes are particularly affected, the frontal cortex often appears most dramatically changed in imaging studies and those changes are most strongly correlated with cognitive deficits in individuals.^{44,45} Although neurons clearly are lost in some regions in some species, it does not appear that normal brain aging involves extensive loss of neurons (as was once thought), not even in the frontal cortex and hippocampus, where the executive and memory functions most affected by aging are regulated. Contemporary stereological studies have established that, in contrast to Alzheimer's disease, there is not extensive and widespread loss of neurons in the normally aging brain, even in individuals with demonstrated cognitive impairment.⁴⁶⁻⁵¹

Aging-related changes appear to involve significant changes in the neuropil and white matter, with alterations in both axons and dendrites. White matter and axonal changes include reductions in the total length of myelinated fibers and apparent loss among fibers of the smallest diameter,⁵²⁻⁵⁴ as well as changes in myelin structure that may compromise neuronal transmission.^{54,55} Several laboratories have reported that neurons in the aging brain undergo a reduction in dendritic length and complexity, as well as a decrease in the density of dendritic spines.^{49,56-60} Changes in dendritic extent are not ubiquitous, however, but rather vary among and within neural regions^{13,61,62} (Figure 1). Loss of dendritic spines may be more widespread.

Significantly, even within the dendritic arbors of individual neurons some components undergo aging-related regression while other components are maintained,⁶³ indicating that changes in dendrites are regulated very focally and may affect specific populations of inputs onto individual neurons.

Since dendritic spines are the primary targets for excitatory synapses,⁶⁴ one would expect loss of dendrites and spines to be associated with a decrease in synaptic number. As for neuronal loss, however, stereological investigations have not supported early reports that there is widespread and dramatic loss of synapses in the aging brain. Stereological analyses of the hippocampus revealed loss of synapses in the dentate gyrus,^{65,66} but the loss appears to involve only subsets of synaptic

inputs.⁶⁷⁻⁶⁹ Synaptic density appears to be largely maintained across aging in other sub-regions of the hippocampus,³² although synapses in CA1 may undergo a small and selective decline.⁷⁰ Analyses of aging-related changes in synaptic number in the cerebral cortex are mixed, even when considering only the frontal cortex, suggesting any changes in synaptic number are restricted in scope and magnitude.⁷¹⁻⁷³ Even in regions in which synapse number is maintained, aging-related changes in dendritic architecture could produce significant rearrangements in the pattern of synaptic inputs on to individual neurons and thereby alter neuronal function.

Aging-related changes in synaptic function

Significant aging-related changes in synaptic transmission and plasticity occur in many neural regions in which there is no apparent loss of synapses or change in their structure, indicating that aging alters aspects of synaptic organization and function that do not have apparent structural correlates.⁷⁴⁻⁷⁷ Many investigations of the synaptic changes that underlie aging-related cognitive deficits have benefited from recognition that individual rodents and non-human primates, like humans, exhibit varying degrees of aging-related cognitive dysfunction. Gallagher and colleagues have demonstrated repeatedly that individual aged rats can be classified as impaired or unimpaired on a number of cognitive tasks, including the hippocampally dependent Morris Water Maze^{32,33,78} and measures of frontal cortical function.⁷⁹ Importantly, impaired individuals show deficits across multiple cognitive domains mediated by different neural regions.⁸⁰ Such evidence that individuals showing less successful aging are impaired on multiple tasks provides confidence that, although many structural changes are restricted to specific sub-regions of the aging brain, some mechanisms contributing to functional changes are represented more globally. Experimentally, the ability to compare cognitively impaired old rats to unimpaired rats of the same age helps investigators differentiate neurobiological changes that contribute to cognitive deficits from aging-related changes that are unrelated to function.

Regardless of whether demonstrated specifically in individuals with demonstrably impaired

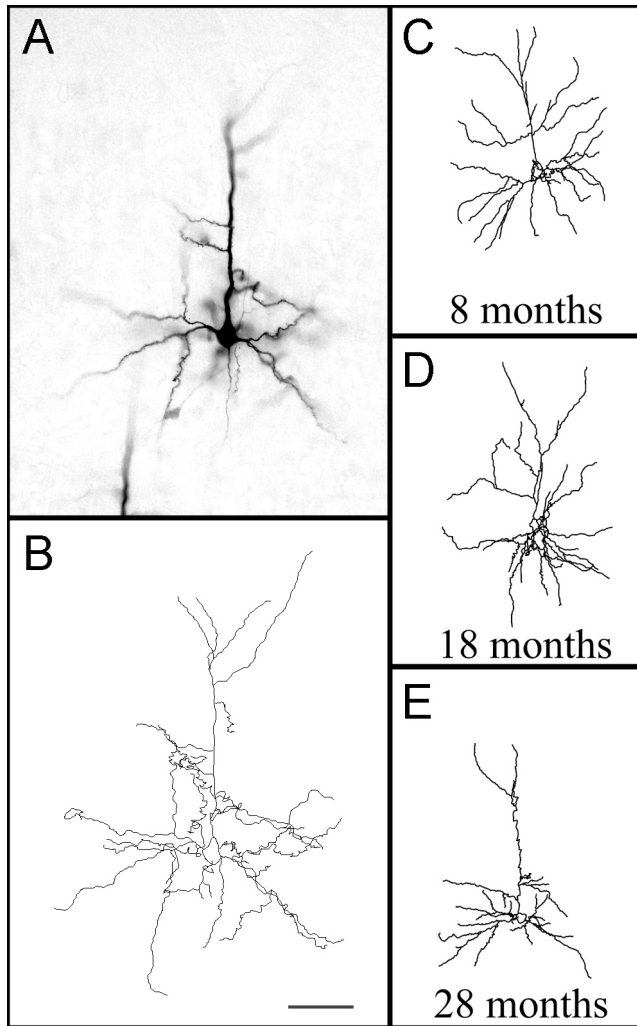


Figure 1. Neuronal labelling and analysis of dendritic extent. A dye-injected, superficial pyramidal neuron is shown (A) with its reconstructed dendritic arbor (B). Scale bar = 40 μm . C, D and E illustrate the dendritic arbors of representative pyramidal neurons from the cingulate cortex of rats at 8, 18 and 28 months of age and demonstrate the decrease in dendritic extent between middle- and old age. Quantitative analysis revealed a 20–25% decrease in dendritic extent for superficial pyramidal neurons, whereas neurons in deeper layers were unchanged.⁶³

cognitive function or simply by comparing animals of different ages, recent studies have provided a wealth of information on how and why synaptic signalling changes with age. Even a cursory overview of the range of synaptic and related intra- and intercellular changes is beyond the scope of this review, but a summary of demonstrated changes in a single neurotransmitter system illustrates the growing sophistication of analyses of synaptic changes.

Glutamate and its receptors mediate the majority of excitatory neurotransmission in the CNS and are critical for neuronal signalling that underlies learning, memory, and other cognitive functions. Glutamate levels have been reported to decline in the aging cerebral cortex and hippocampus, but that change may be more related to metabolic activity than to synaptic transmission.⁸¹ The most significant aging-related changes in glutamate signalling appear to arise

from many and varied changes in glutamate receptors. Glutamate signals through multiple receptors of two general classes, ionotropic receptors that form membrane-spanning ion channels and signal through ion fluxes, and metabotropic receptors that signal through intracellular second messenger systems. Each receptor type in each class comprises multiple sub-units.^{82,83} Changes in glutamate signaling through the ionotropic N-methyl D-aspartate (NMDA) type receptor appear to underlie many changes in neural function and for detailed descriptions of aging-related changes in glutamate and NMDA signalling readers are directed to recent focused reviews.^{59,81,82,84–88} In brief, NMDA receptors are expressed at high levels in the cerebral cortex and hippocampus and play critical roles in learning and memory and in many plasticity processes. Selective blockade of NMDA receptors impairs performance on memory tasks that are affected by aging. Several studies have demonstrated a correlation between lower densities of NMDA receptor-binding and poor memory performance. The density of NMDA receptors is reduced in some areas of the aging brain, but such changes are region-specific. In addition to the many changes in NMDA receptor function, metabotropic glutamate receptors also undergo aging-related changes in expression and ligand-binding that contribute to changes in their function.^{89–91}

For both classes of glutamate receptors, changes in the expression and balance of specific receptor subunits and alterations in the insertion of receptors in the post-synaptic membrane alter function even in the absence of changes in overall numbers of receptors. In addition, intracellular pathways that are coupled to the receptors undergo significant aging-related changes that blunt or alter the fidelity of signalling.^{87,92,93} Thus, even if one considers only the receptors for a single neurotransmitter, one finds abundant substrates for physiological and pathophysiological regulation that influences neural processing and cognition during aging.

Fundamental mechanisms of brain aging – oxidative stress and inflammation

The breadth of aging-related neurobiological changes – including alterations in many different neuron types and changes in some structural

properties and many inter- and intracellular signalling pathways – raises the question of whether a small number of cellular and molecular regulators could underlie the variety of structural and functional changes that result in cognitive deficits. Accumulating evidence indicates that two important and related changes in the microenvironment within the brain contribute to many of the cellular and intercellular changes that occur with aging: progressive increases in oxidative stress and neuroinflammation.

Oxidative damage increases with age,^{94–97} due to increased generation of reactive oxygen species (ROS), decreased anti-oxidant activity, and/or deficient mechanisms to prevent or repair cell and tissue damage. It is not clear whether anti-oxidant mechanisms and repair mechanisms are significantly impaired with age in all tissues,⁹⁸ but the levels of key antioxidant enzymes clearly decline in some regions of the brain,^{99–101} possibly contributing to increased oxidative damage.^{96,102–104} There is evidence for a close association between oxidative stress and aging-related functional decline in some tissues,⁹⁵ with oxidative stress inducing greater DNA and protein damage in older animals.^{105,106} The increase in damaged proteins appears to be regionally specific, with the hippocampus exhibiting a twofold greater increase in oxidative damage than cortical structures.⁹⁵ Importantly, increased oxidative stress has been associated directly with impairments in motor and cognitive performance.^{95,107} In addition, interventions that reduce oxidative damage (e.g. moderate caloric restriction, high antioxidant diets, or spin trapping agents) prevent or reverse some behavioral deficits observed in aged animals.^{96,108–110}

It is reasonable to expect that increased oxidative stress would affect cellular structure and a wide variety of cellular and intercellular processes, since essentially all macromolecules in the brain – proteins, lipids and nucleic acids – are subject to oxidative damage. Moreover, recent studies have begun to reveal additional and dynamic effects of the imbalance between ROS and antioxidant defenses in the aged brain. Oxidative conditions affect a variety of redox-sensitive signalling processes, including the insulin-receptor signalling pathways. These pathways influence neurodegenerative processes and normal brain aging,^{111,112} and may even influence lifespan.¹¹³ Thus, oxidative stress in the aging brain appears to

affect multiple and complex regulatory networks within the brain.

Much of the aging-related increase in oxidative stress is likely to be an unavoidable result of metabolic processes, but ROS also are produced as part of neuroinflammatory processes that increase with age. Accumulating evidence suggests a loss of balance between pro- and anti-inflammatory signals that contributes to loss of neural function. Elevations in pro-inflammatory markers commonly are found in the CNS of healthy older people and laboratory animals.^{114–118} Indeed, gene array studies screening for neural genes with altered expression in the aging brain found that a large percentage of the genes that showed an age-related alteration in expression were involved in the inflammatory response or in regulating oxidative stress.^{119,120} More targeted studies demonstrated an age-dependent increase in transcription of the pro-inflammatory genes *TNF- α* , *IL-1 β* , and *MCP-1* in the hippocampus of old versus young adult mice.¹²¹ Importantly, altered regulation of inflammatory gene expression in older rodents was associated with age-related cognitive impairment; in cognitively impaired old rats expression was altered but in unimpaired old rats expression was similar to that in young adults.¹⁰⁷ The gene-expression studies are supported by evidence that basal levels of pro-inflammatory proteins are increased in the brains of old rodents compared to young.^{122–124}

In addition to these molecular and biochemical studies, cellular effects of aging on inflammatory processes in the brain are demonstrated by changes in microglia, the cells that initiate and direct the inflammatory response in the CNS. In the absence of pro-inflammatory stimuli, microglia are characterized morphologically by the presence of long, thin, and ramified cellular processes and often are referred to as ‘resting’. Resting is a poor descriptor, however, since these microglia actively monitor their microenvironment^{125–127} via a wide variety of cell-surface receptors.^{128–131} Microglia respond to chemical, physical, infectious, and other factors that signal or may cause damage with morphological changes, including partial retraction and thickening of the cellular processes and rounding of the cell body, and functional changes, including migration, proliferation, and the production and secretion of pro-inflammatory proteins.¹³² In the adult brain, the pro-inflammatory cytokines produced by activated microglia influence neurogenesis,^{133–135} long-term

potentiation (LTP),¹³⁶ and cognition.^{137,138} The importance of such changes for aging-related cognitive decline is supported by demonstrations that age-related impairments in LTP and cognition correlate with increased levels of *IL-1 β* , increased numbers of activated microglia, and decreased levels of the anti-inflammatory cytokines *IL4* and *IL10*.^{136,139–141}

Immunohistochemical studies have demonstrated an increase in the number of activated microglia in the CNS of healthy, older rodents,^{142–145} non-human primates,¹⁴⁶ and humans.^{147,148} Aging apparently alters the functional state of microglia but not their number,^{145,149} since significant functional changes occur in individual cells with little change in the size of the population. The increase in microglial activation with age is dramatic and easily demonstrated with immunomarkers that target the phagocytic nature of activated microglia or changes in cell surface receptors (Figure 2). It remains a major challenge, however, to characterize specifically the functional changes in microglia in specific regions of the aging brain. This is a critical issue in light of accumulating evidence that, depending on their functional state and the factors acting upon them, microglia may have trophic actions that support neuronal survival and function,^{127,150–154} as well as deleterious effects on neurons and other cells. Transgenic mice with fluorescently tagged microglia offer significant promise for detailed analyses of cellular and molecular changes in activated microglia,¹⁵⁵ permitting comparisons among neural regions with greater and lesser degrees of aging-related functional changes.

Ameliorating aging-related cognitive decline

Evidence that oxidative stress and inflammatory processes underlie many aging-related functional changes in the brain makes these processes attractive targets for therapies to prevent or reverse aging-related cognitive decline. In animal models, diets that are rich in anti-oxidants have proven effective in ameliorating some cognitive deficits in older animals (see recent reviews^{110,152,156–159}). Anti-inflammatory treatments can decrease the number of activated microglia in the brain and also partially restore the age-dependent impairment in LTP¹³⁶ and spatial memory performance.¹³⁷ Similarly, inhibition of the enzyme responsible

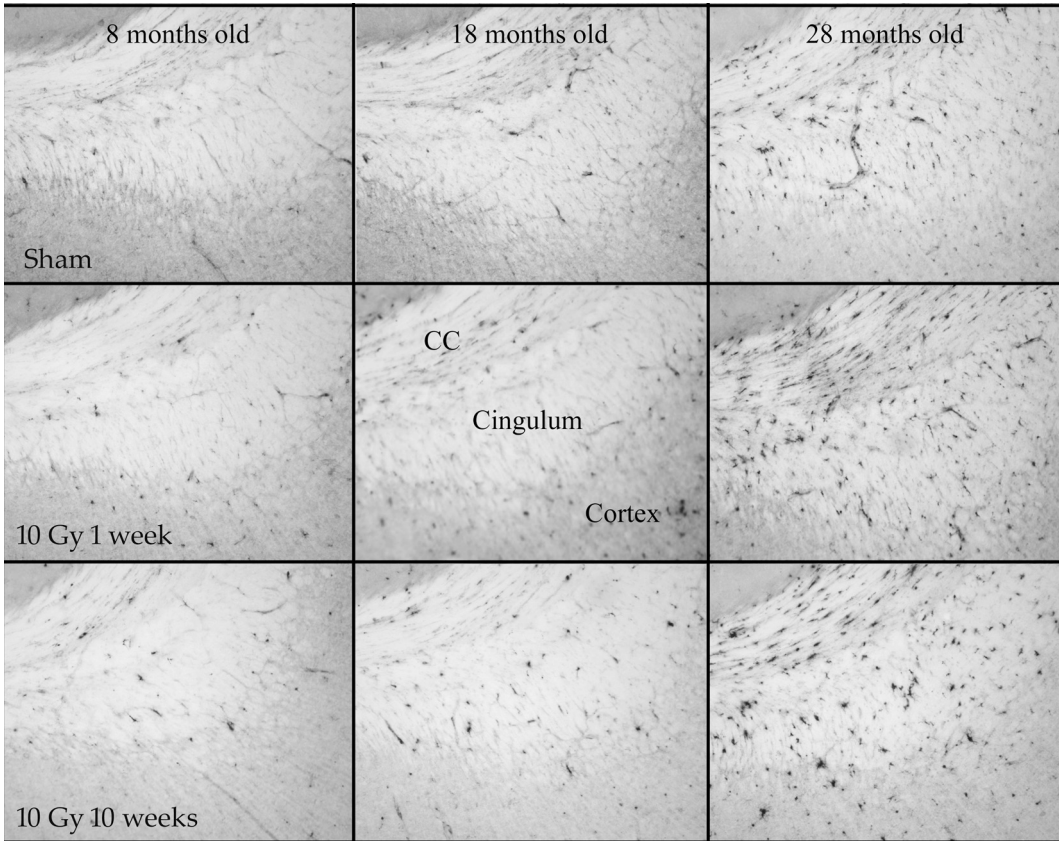


Figure 2. Aging-related increase in activated microglia and in the inflammatory response to brain irradiation. Activated microglia can be identified using the ED1 monoclonal antibody, which recognizes a lysosomal protein (CD68) that is upregulated when microglia become activated. ED1⁺ microglia are rare in the brain of a young control rat but increase in abundance with age (top row). A single 10 Gy dose of whole-brain irradiation produces only a modest activation of microglia in the young brain (left column) but a much greater and sustained activation in middle-aged and old rats (middle and right columns). ED1 labelling is shown in the corpus callosum (CC), cingulum and deep cerebral cortex, but similar changes are seen throughout the brain.¹⁴⁵

for activating IL-1 β improves age-related memory dysfunction.¹³⁵

Although such animal studies indicate that antioxidant and anti-inflammatory therapies might limit aging-related cognitive deficits, evidence that oxidative damage and pro-inflammatory conditions in the human brain accumulate over many years raise the critical question of whether it would be necessary to begin therapies early in life to derive benefits. Animal studies demonstrate that supplementation with dietary antioxidants for as little as 2–4 months can reduce normal aging-related cognitive deficits in rodents.^{160,161} As yet, however, interventional studies of antioxidant supplementation in humans have largely failed to demonstrate benefits for

cognitive function,^{162–164} despite observational evidence of an association between nutritional intake of antioxidants and cognitive function in healthy elderly individuals.^{165,166} Presently, there may be more translational relevance and clinical potential in studies demonstrating that antioxidant and anti-inflammatory treatments are efficacious in preventing or decreasing the effects of potentially damaging challenges, to which the aged brain is particularly vulnerable.^{167,168}

Increased vulnerability to damage in the aging brain

Clinical experience and anecdotal and experimental evidence indicate that older individuals

have greater difficulty recovering from events that challenge the homeostatic mechanisms of the brain. Significantly, such challenges typically induce oxidative stress and/or inflammation, and many are common among older individuals. For example, postoperative cognitive dysfunction is more common in older than in younger adult patients, even in the absence of adverse intraoperative events.^{169–171} As this problem has become more recognized clinically, experimental studies are beginning to investigate the underlying mechanism^{172,173}; oxidative stress and inflammatory effects almost certainly may play an important role. Clinical and experimental studies of stroke, traumatic brain injury, axotomy, and whole-brain irradiation all support the hypothesis that aging impacts the intensity and duration of brain inflammation and microglial activation following challenges^{145,147,174,175} (Figure 2). Functionally, old age alters the duration of radiation-induced cognitive deficits in rodents,^{176,177} and clinical studies have identified increased age at the time of irradiation to be a risk factor for development and severity of radiation-induced cognitive side-effects.^{178–181}

Such differences between young adult and older brains indicate that the neural effects of potentially damaging challenges may be quantitatively greater, and even mechanistically different, in middle-aged and elderly patients, presumably because older brains are in a chronic state of oxidative stress and inflammation (discussed in Schindler *et al*¹⁴⁵). Microglia often exhibit an increased response to an acute inflammatory event when the event was preceded by a previous inflammatory stimulus; in the CNS such microglial ‘priming’ may even involve interactions between the systemic and parenchymal immune systems.^{182–186} Microglia in the aged brain may be primed to produce an exaggerated response to any pro-inflammatory and pro-oxidative stimulus, such that a challenge that produces no significant functional effects in young individuals elicits a feed-forward and progressive oxidative stress and neuroinflammatory response that impairs neuronal function in elderly individuals. Thus, even if treatment with antioxidant and anti-inflammatory factors does not improve basal cognitive function in the aging and aged, it may provide protection from the additional cognitive deficits that often result from the inevitable neural challenges that are associated with growing older.

Ongoing and future studies

As the survey above demonstrates, there has been tremendous progress in (1) elucidating elements of cellular structure and intercellular signalling that are compromised in the aging brain and (2) developing evidence that increased oxidative stress and neuroinflammation are critical contributors to the neurobiological changes that result in cognitive dysfunction. A primary focus now and in future studies is revealing the mechanistic pathways through which oxidative stress and inflammation alter neurotransmitter receptors, synapses, and neuronal processes.¹⁸⁷ These mechanistic links are beginning to be revealed. For example, tumor necrosis factor- α (TNF- α) interleukin 1-beta and interleukin 18 all inhibit LTP, and TNF- α alters expression of metabotropic glutamate receptors and potentiates glutamate excitotoxicity.^{188,189} TNF- α also inhibits neurite outgrowth and thereby may limit recovery following neuronal damage.¹⁹⁰ Interferon- α suppresses NMDA signalling¹⁹¹ and interleukin-6 produces subunit-specific reductions in metabotropic glutamate receptors.¹⁹² Microglia can influence a variety of synaptic properties¹⁹³ and, under some conditions, activated microglia can even strip synapses from neurons.¹⁹⁴

The continued revelation of the influence of oxidative stress and inflammation on the structural and functional mediators of aging-related cognitive decline should produce new targets for developing therapies to ameliorate aging-related cognitive dysfunction and more refined assays to test those therapies and promote their translation to the clinic.

Acknowledgements

The authors’ research on brain aging is supported by National Institutes of Health grant AG 11370 (DRP). The authors thank Jen Sousa for her assistance in the preparation of the manuscript.

References

- 1 Gliskey EL. Changes in cognitive function in human aging. In: Riddle DR ed. *Brain aging: models, methods and mechanisms*. Boca Raton: CRC Press, 2007: 3–20.
- 2 Verhaeghen P, Cerella J. Aging, executive control, and attention: a review of meta-analyses. *Neurosci Biobehav Rev* 2002; 26: 849–57.

- 3 Thornton WJ, Raz N. Aging and the role of working memory resources in visuospatial attention. *Neuropsychol Dev Cogn B Aging Neuropsychol Cogn* 2006; **13**: 36–61.
- 4 McDowd JM, Shaw RJ. Human memory. In: Craik FIM, Salthouse TA eds. *The handbook of aging and cognition*, second edition. Mahwah, NJ: Erlbaum, 2000: 221–92.
- 5 Verhaeghen P, Basak C. Ageing and switching of the focus of attention in working memory: results from a modified N-back task. *Q J Exp Psychol A* 2005; **58**: 134–54.
- 6 Hogan MJ, Kelly CA, Craik FI. The effects of attention-switching on encoding and retrieval of words in younger and older adults. *Exp Aging Res* 2006; **32**: 153–83.
- 7 Hillman CH, Erickson KI, Kramer AF. Be smart, exercise your heart. Exercise effects on brain cognition. *Nature Rev Neurosci* 2008; **9**: 58–65
- 8 Kester JD, Benjamin AS, Castel AD, Craik FIM. Memory in elderly people. In: Baddeley AD, Kopelman MD, Wilson BA eds. *The handbook of memory disorders*. West Sussex: Wiley, 2002: 543–68.
- 9 Luszcz MA, Bryan J. Toward understanding age-related memory-loss in late adulthood. *Gerontology* 1999; **45**: 2–9.
- 10 Grady CL, Craik FI. Changes in memory processing with age. *Curr Opin Neurobiol* 2000; **10**: 224–31.
- 11 de Fockert JW. Keeping priorities: the role of working memory and selective attention in cognitive aging. *Sci Aging Knowledge Environ* 2005; **44**: 34.
- 12 Buckner RL. Memory and executive function in aging and AD: multiple factors that cause decline and reserve factors that compensate. *Neuron* 2004; **44**: 195–208.
- 13 Burke SN, Barnes CA. Neural plasticity in the ageing brain. *Nat Rev Neurosci* 2006; **7**: 30–40.
- 14 Tanji J, Hoshi E. Role of the lateral prefrontal cortex in executive behavioral control. *Physiol Rev* 2008; **88**: 37–57.
- 15 West RL. An application of prefrontal cortex function theory to cognitive aging. *Psychol Bull* 1996; **120**: 272–92.
- 16 Resnick SM, Pham DL, Kraut MA, Zonderman AB, Davatzikos C. Longitudinal magnetic resonance imaging studies of older adults: a shrinking brain. *J Neurosci* 2003; **23**: 3295–301.
- 17 Hedden T. Imaging cognition in the aging human brain. In: Riddle DR ed. *Brain aging: models, methods and mechanisms*. Boca Raton: CRC Press, 2007: 251–78.
- 18 Raz N, Rodrigue KM, Haacke EM. Brain aging and its modifiers: insights from in vivo neuromorphometry and susceptibility weighted imaging. *Ann N Y Acad Sci* 2007; **1097**: 84–93.
- 19 Prvulovic D, Van de Ven V, Sack AT, Maurer K, Linden DE. Functional activation imaging in aging and dementia. *Psychiatry Res* 2005 Nov 30; **140**: 97–113.
- 20 Rajah MN, D'Esposito M. Region-specific changes in prefrontal function with age: a review of PET and fMRI studies on working and episodic memory. *Brain* 2005; **128**(Pt 9): 1964–83.
- 21 Kessler RM. Imaging methods for evaluating brain function in man. *Neurobiol Aging* 2003; **24** (suppl 1): 21–35; discussion S37–39.
- 22 Meltzer CC, Becker JT, Price JC, Moses-Kolko E. Positron emission tomography imaging of the aging brain. *Neuroimaging Clin N Am* 2003; **13**: 759–67.
- 23 Charlton RA, McIntyre DJ, Howe FA, Morris RG, Markus HS. The relationship between white matter brain metabolites and cognition in normal aging: the GENIE study. *Brain Res* 2007; **1164**: 108–16.
- 24 McIntyre DJ, Charlton RA, Markus HS, Howe FA. Long and short echo-time proton magnetic resonance spectroscopic imaging of the healthy aging brain. *J Magn Reson Imaging* 2007; **26**: 1596–606.
- 25 Petrella JR, Mattay VS, Doraiswamy PM. Imaging genetics of brain longevity and mental wellness: the next frontier? *Radiology* 2008; **246**: 20–32.
- 26 Zahr NM, Mayer D, Pfefferbaum A, Sullivan EV. Low Striatal glutamate levels underlie cognitive decline in the elderly: Evidence from in vivo molecular spectroscopy. *Cereb Cortex* 2008 Jan 29; [Epub ahead of print].
- 27 Nadon NL. Of mice and monkeys: National Institute on Aging resources supporting the use of animal models in biogerontology research. *J Gerontol A Biol Sci Med Sci* 2006; **61**: 813–15.
- 28 Dias R, Robbins TW, Roberts AC. Primate analogue of the Wisconsin Card Sorting Test: effects of excitotoxic lesions of the prefrontal cortex in the marmoset. *Behav Neurosci* 1996; **110**: 872–86.
- 29 Dias R, Robbins TW, Roberts AC. Dissociation in prefrontal cortex of affective and attentional shifts. *Nature* 1996; **380**(6569): 69–72.
- 30 Herndon JG, Lacreuse A. The Rhesus Monkey Model as a heuristic resource in cognitive aging research. In: Erwin JM, Hof PR ed. *Aging in nonhuman primates. Interdiscipl Top Gerontol Basel*. Karger, 2002; **31**: 178–195.
- 31 Moss MB, Moore TL, Schettler SP, Killiany R, Rosene D. Successful vs. unsuccessful aging in the

- rhesus monkey. In: Riddle DR ed. *Brain aging: models, methods and mechanisms*. Boca Raton: CRC Press, 2007: 21–38.
- 32 Geinisman Y, Ganeshina O, Yoshida R, Berry RW, Disterhoft JF, Gallagher M. Aging, spatial learning, and total synapse number in the rat CA1 stratum radiatum. *Neurobiol Aging* 2004; **25**: 407–16.
- 33 Nicholson DA, Yoshida R, Berry RW, Gallagher M, Geinisman Y. Reduction in size of perforated postsynaptic densities in hippocampal axospinous synapses and age-related spatial learning impairments. *J Neurosci* 2004; **24**: 7648–53.
- 34 Driscoll I, Sutherland RJ. The aging hippocampus: navigating between rat and human experiments. *Rev Neurosci* 2005; **16**: 87–121.
- 35 Lee HK, Min SS, Gallagher M, Kirkwood A. NMDA receptor-independent long-term depression correlates with successful aging in rats. *Nat Neurosci* 2005; **8**: 1657–59.
- 36 McGaughy J, Ross RS, Eichenbaum H. Noradrenergic, but not cholinergic, deafferentation of prefrontal cortex impairs attentional set-shifting. *Neuroscience* 2008 Feb 19; [Epub ahead of print].
- 37 Barense MD, Fox MT, Baxter MG. Aged rats are impaired on an attentional set-shifting task sensitive to medial frontal cortex damage in young rats. *Learn Mem* 2002; **9**: 191–201.
- 38 Brown VJ, Bowman EM. Rodent models of prefrontal cortical function. *Trends Neurosci* 2002; **25**: 340–43.
- 39 Andersen JK. Genetically engineered mice and their use in aging research. *Mol Biotechnol* 2001; **19**: 45–57.
- 40 Martin GM. Genetic engineering of mice to test the oxidative damage theory of aging. *Ann N Y Acad Sci*. 2005; **1055**: 26–34.
- 41 Bartke A. New findings in gene knockout, mutant and transgenic mice. *Exp Gerontol* 2008; **43**: 11–14.
- 42 Peters A, Morrison JH, Rosene DL, Hyman BT. Feature article: are neurons lost from the primate cerebral cortex during normal aging? *Cereb Cortex* 1998; **8**: 295–00.
- 43 Jernigan TL, Archibald SL, Fennema-Notestine C *et al*. Effects of age on tissues and regions of the cerebrum and cerebellum. *Neurobiol Aging* 2001; **22**: 581–94.
- 44 Van Petten C, Plante E, Davidson PS, Kuo TY, Bajuscak L, Glisky EL. Memory and executive function in older adults: relationships with temporal and prefrontal gray matter volumes and white matter hyperintensities. *Neuropsychologia* 2004; **42**: 1313–35.
- 45 Allen JS, Bruss J, Brown CK, Damasio H. Normal neuroanatomical variation due to age: the major lobes and a parcellation of the temporal region. *Neurobiol Aging* 2005; **26**: 1245–60.
- 46 West MJ. Regionally specific loss of neurons in the aging human hippocampus. *Neurobiol Aging* 1993; **14**: 287–93.
- 47 Rapp PR, Gallagher M. Preserved neuron number in the hippocampus of aged rats with spatial learning deficits. *Proc Natl Acad Sci USA* 1996; **93**: 9926–30.
- 48 Rasmussen T, Schliemann T, Sorensen JC, Zimmer J, West MJ. Memory-impaired aged rats: no loss of principal hippocampal and subicular neurons. *Neurobiol Aging* 1996; **17**: 143–47.
- 49 Hof PR, Morrison JH. The aging brain: morphomolecular senescence of cortical circuits. *Trends Neurosci* 2004; **27**: 607–13.
- 50 Dickstein DL, Kabaso D, Rocher AB, Luebke JI, Wearne SL, Hof PR. Changes in the structural complexity of the aged brain. *Aging Cell* 2007; **6**: 275–84.
- 51 Morrison JH, Hof PR. Life and death of neurons in the aging cerebral cortex. *Int Rev Neurobiol* 2007; **81**: 41–57.
- 52 Tang Y, Nyengaard JR, Pakkenberg B, Gundersen HJ. Age-induced white-matter changes in the human brain: a stereological investigation. *Neurobiol Aging* 1997; **18**: 609–15.
- 53 Marner L, Nyengaard JR, Tang Y, Pakkenberg B. Marked loss of myelinated nerve fibers in the human brain with age. *J Comp Neurol* 2003; **462**: 144–52.
- 54 Hinman JD, Abraham CR. What's behind the decline? The role of white matter in brain aging. *Neurochem Res* 2007; **32**: 2023–31.
- 55 Peters A. The effects of normal aging on myelin and nerve fibers: a review. *J Neurocytol* 2002; **31**: 581–93.
- 56 Nimchinsky EA, Sabatini BL, Svoboda K. Structure and function of dendritic spines. *Annu Rev Physiol* 2002; **64**: 313–53.
- 57 Halpain S, Spencer K, Graber S. Dynamics and pathology of dendritic spines. *Prog Brain Res* 2005; **147**: 29–37.
- 58 von Bohlen and Halbach O, Zacher C, Gass P, Unsicker K. Age-related alterations in hippocampal spines and deficiencies in spatial memory in mice. *J Neurosci Res* 2006; **83**: 525–31.
- 59 Morrison JH, Hof PR. Selective vulnerability of corticocortical and hippocampal circuits in aging and Alzheimer's disease. *Prog Brain Res* 2002; **136**: 467–86.
- 60 Wallace M, Frankfurt M, Arellanos A, Inagaki T, Luine V. Impaired recognition memory and decreased prefrontal cortex spine density in aged female rats. *Ann N Y Acad Sci* 2007; **1097**: 54–57

- 61 Coleman PD, Flood DG. Net dendritic stability of layer II pyramidal neurons in F344 rat entorhinal cortex from 12 to 37 months. *Neurobiol Aging* 1991; **12**: 535–41.
- 62 Hanks SD, Flood DG. Region-specific stability of dendritic extent in normal human aging and regression in Alzheimer's disease. I. CA1 of hippocampus. *Brain Res* 1991; **540**: 63–82.
- 63 Grill JD, Riddle DR. Age-related and laminar-specific dendritic changes in the medial frontal cortex of the rat. *Brain Res* 2002; **937**: 8–21.
- 64 Alvarez VA, Sabatini BL. Anatomical and physiological plasticity of dendritic spines. *Annu Rev Neurosci* 2007; **30**: 79–97.
- 65 Geinisman Y, de Toledo-Morrell L, Morrell F, Persina IS, Rossi M. Age-related loss of axospinous synapses formed by two afferent systems in the rat dentate gyrus as revealed by the unbiased stereological disector technique. *Hippocampus* 1992; **2**: 437–44.
- 66 Rutten BP, Van Der Kolk NM, Schafer S *et al*. Age-related loss of synaptophysin immunoreactive presynaptic boutons within the hippocampus of APP751SL, PS1M146L, and APP751SL/PS1M146L transgenic mice. *Am J Pathol* 2005; **167**: 161–73.
- 67 Tigges J, Herndon JG, Rosene DL. Mild age-related changes in the dentate gyrus of adult rhesus monkeys. *Acta Anat (Basel)* 1995; **153**: 39–48.
- 68 Tigges J, Herndon JG, Rosene DL. Preservation into old age of synaptic number and size in the supragranular layer of the dentate gyrus in rhesus monkeys. *Acta Anat (Basel)* 1996; **157**: 63–72.
- 69 Newton IG, Forbes ME, Linville MC *et al*. Effects of aging and caloric restriction on dentate gyrus synapses and glutamate receptor subunits. *Neurobiol Aging* 2007 Apr 10; [Epub ahead of print]
- 70 Shi L, Linville MC, Tucker EW, Sonntag WE, Brunso-Bechtold JK. Differential effects of aging and insulin-like growth factor-1 on synapses in CA1 of rat hippocampus. *Cereb Cortex* 2005; **15**: 571–77.
- 71 Scheff SW, Price DA, Sparks DL. Quantitative assessment of possible age-related change in synaptic numbers in the human frontal cortex. *Neurobiol Aging* 2001; **22**: 355–65.
- 72 Shi L, Pang H, Linville MC, Bartley AN, Argenta AE, Brunso-Bechtold JK. Maintenance of inhibitory interneurons and boutons in sensorimotor cortex between middle and old age in Fischer 344 X Brown Norway rats. *J Chem Neuroanat* 2006; **32**: 46–53.
- 73 Peters A, Sethares C, Luebke JI. Synapses are lost during aging in the primate prefrontal cortex. *Neuroscience* 2007 Jul 17; [Epub ahead of print]
- 74 Barnes CA. Long-term potentiation and the ageing brain. *Philos Trans R Soc Lond B Biol Sci* 2003; **358**(1432): 765–72.
- 75 Gallagher M. Aging and hippocampal/cortical circuits in rodents. *Alzheimer Dis Assoc Disord* 2003; **17** (suppl 2): S45–7.
- 76 Rosenzweig ES, Barnes CA. Impact of aging on hippocampal function: plasticity, network dynamics, and cognition. *Prog Neurobiol* 2003; **69**: 143–79.
- 77 Wilson IA, Gallagher M, Eichenbaum H, Tanila H. Neurocognitive aging: prior memories hinder new hippocampal encoding. *Trends Neurosci* 2006; **29**: 662–70.
- 78 Zhang HY, Watson ML, Gallagher M, Nicolle MM. Muscarinic receptor-mediated GTP-Eu binding in the hippocampus and prefrontal cortex is correlated with spatial memory impairment in aged rats. *Neurobiol Aging* 2007; **28**: 619–26.
- 79 Schoenbaum G, Setlow B, Saddoris MP, Gallagher M. Encoding changes in orbitofrontal cortex in reversal-impaired aged rats. *J Neurophysiol* 2006; **95**: 1509–17.
- 80 LaSarge CL, Montgomery KS, Tucker C *et al*. Deficits across multiple cognitive domains in a subset of aged Fischer 344 rats. *Neurobiol Aging* 2007; **28**: 928–36.
- 81 Segovia G, Porras A, Del Arco A, Mora F. Glutamatergic neurotransmission in aging: a critical perspective. *Mech Ageing Dev* 2001; **122**: 1–29.
- 82 Michaelis EK. Molecular biology of glutamate receptors in the central nervous system and their role in excitotoxicity, oxidative stress and aging. *Prog Neurobiol* 1998; **54**: 369–415.
- 83 Dingledine R, Borges K, Bowie D, Traynelis SF. The glutamate receptor ion channels. *Pharmacol Rev* 1999; **51**: 7–61.
- 84 Magnusson KR. The aging of the NMDA receptor complex. *Front Biosci* 1998; **3**: 70–80.
- 85 Yamada K, Nabeshima T. Changes in NMDA receptor/nitric oxide signaling pathway in the brain with aging. *Microsc Res Tech* 1998; **43**: 6X–74.
- 86 Adams MM, Morrison JH. Estrogen and the aging hippocampal synapse. *Cereb Cortex* 2003; **13**: 1271–75.
- 87 Foster TC. Calcium homeostasis and modulation of synaptic plasticity in the aged brain. *Aging Cell* 2007; **6**: 319–25.
- 88 Shi L, Adams M, Brunso-Bechtold JK. Subtle alterations in glutamatergic receptors underlie the aging-related decline in hippocampal function. In: Riddle, DR ed. *Brain aging: models, methods and*

- mechanisms*. Boca Raton: CRC Press, 2007: 189–212.
- 89 Simonyi A, Miller LA, Sun GY. Region-specific decline in the expression of metabotropic glutamate receptor 7 mRNA in rat brain during aging. *Brain Res Mol Brain Res* 2000; **82**: 101–06.
 - 90 Simonyi A, Ngomba RT, Storto M *et al*. Expression of groups I and II metabotropic glutamate receptors in the rat brain during aging. *Brain Res* 2005; **1043**: 95–106.
 - 91 Hedberg TG, Velísková J, Sperber EF, Nunes ML, Moshé SL. Age-related differences in NMDA/metabotropic glutamate receptor-binding in rat substantia nigra. *Int J Dev Neurosci* 2003; **21**: 95–103.
 - 92 Nicolle MM, Colombo PJ, Gallagher M, McKinney M. Metabotropic glutamate receptor-mediated hippocampal phosphoinositide turnover is blunted in spatial learning-impaired aged rats. *J Neurosci* 1999; **19**: 9604–10.
 - 93 Domenici MR, Pintor A, Potenza RL *et al*. Metabotropic glutamate receptor 5 (mGluR5)-mediated phosphoinositide hydrolysis and NMDA-potentiating effects are blunted in the striatum of aged rats: a possible additional mechanism in striatal senescence. *Eur J Neurosci* 2003; **17**: 2047–55.
 - 94 Sohal RS, Agarwal S, Sohal BH. Oxidative stress and aging in the Mongolian gerbil (*Meriones unguiculatus*). *Mech Ageing Dev* 1995; **81**: 15–25.
 - 95 Forster MJ, Dubey A, Dawson KM, Stutts WA, Lal H, Sohal RS. Age-related losses of cognitive function and motor skills in mice are associated with oxidative protein damage in the brain. *Proc Natl Acad Sci U S A* 1996; **93**: 4765–69.
 - 96 Sohal RS, Weindruch R. Oxidative stress, caloric restriction, and aging. *Science* 1996; **273**: 59–63.
 - 97 Hamilton ML, Van Remmen H, Drake JA *et al*. Does oxidative damage to DNA increase with age? *Proc Natl Acad Sci U S A* 2001; **98**: 10469–74.
 - 98 Agarwal S, Sohal RS. Relationship between susceptibility to protein oxidation, aging, and maximum lifespan potential of different species. *Exp Gerontol* 1996; **31**: 365–72.
 - 99 Chen TS, Richie JP, Jr., Lang CA. The effect of aging on glutathione and cysteine levels in different regions of the mouse brain. *Proc Soc Exp Biol Med* 1989; **190**: 399–402.
 - 100 Rao G, Xia E, Richardson A. Effect of age on the expression of antioxidant enzymes in male Fischer F344 rats. *Mech Ageing Dev* 1990; **53**: 49–60.
 - 101 Semsei I, Rao G, Richardson A. Expression of superoxide dismutase and catalase in rat brain as a function of age. *Mech Ageing Dev* 1991; **58**: 13–19.
 - 102 LeBel CP, Bondy SC. Oxidative damage and cerebral aging. *Prog Neurobiol* 1992; **38**: 601–09.
 - 103 Butterfield DA, Howard B, Yatin S *et al*. Elevated oxidative stress in models of normal brain aging and Alzheimer's disease. *Life Sci* 1999; **65**: 1883–92.
 - 104 Liu J, Mori A. Stress, aging, and brain oxidative damage. *Neurochem Res* 1999; **24**: 1479–97.
 - 105 Sohal RS, Agarwal S, Candas M, Forster MJ, Lal H. Effect of age and caloric restriction on DNA oxidative damage in different tissues of C57BL/6 mice. *Mech Ageing Dev* 1994; **76**: 215–24.
 - 106 Sohal RS, Ku HH, Agarwal S, Forster MJ, Lal H. Oxidative damage, mitochondrial oxidant generation and antioxidant defenses during aging and in response to food restriction in the mouse. *Mech Ageing Dev* 1994; **74**: 121–33.
 - 107 Nicolle MM, Gonzalez J, Sugaya K *et al*. Signatures of hippocampal oxidative stress in aged spatial learning-impaired rodents. *Neuroscience* 2001; **107**: 415–31.
 - 108 Carney JM, Starke-Reed PE, Oliver CN *et al*. Reversal of age-related increase in brain protein oxidation, decrease in enzyme activity, and loss in temporal and spatial memory by chronic administration of the spin-trapping compound N-tert-butyl-alpha-phenylnitron. *Proc Natl Acad Sci USA* 1991; **88**: 3633–36.
 - 109 Wang Y, Chang CF, Chou J *et al*. Dietary supplementation with blueberries, spinach, or spirulina reduces ischemic brain damage. *Exp Neurol* 2005; **193**: 75–84.
 - 110 Joseph JA, Shukitt-Hale B, Lau FC. Fruit polyphenols and their effects on neuronal signaling and behavior in senescence. *Ann N Y Acad Sci* 2007; **100**: 470–85.
 - 111 Calabrese V, Guagliano E, Sapienza M *et al*. Redox regulation of cellular stress response in aging and neurodegenerative disorders: role of vitamins. *Neurochem Res* 2007; **32**: 757–73.
 - 112 Dröge W, Schipper HM. Oxidative stress and aberrant signaling in aging and cognitive decline. *Ageing Cell* 2007; **6**: 361–70.
 - 113 Taguchi A, White MF. Insulin-like signalling, nutrient homeostasis, and life span. *Annu Rev Physiol* 2008; **70**: 191–212.
 - 114 Prolla TA, Mattson MP. Molecular mechanisms of brain aging and neurodegenerative disorders: lessons from dietary restriction. *Trends Neurosci* 2001; **24**: S21–S31.
 - 115 Ye SM, Johnson RW. An age-related decline in interleukin-10 may contribute to the increased expression of interleukin-6 in brain of aged mice. *Neuroimmunomodulation* 2001; **9**: 183–92.
 - 116 Sharman KG, Sharman EH, Yang E, Bondy SC. Dietary melatonin selectively reverses age-related changes in cortical cytokine mRNA levels, and

- their responses to an inflammatory stimulus. *Neurobiol Aging* 2002; **23**: 633–38.
- 117 Weaver JD, Huang MH, Albert M, Harris T, Rowe JW, Seeman TE. Interleukin-6 and risk of cognitive decline: MacArthur studies of successful aging. *Neurology* 2002; **59**: 371–78.
- 118 Bodles AM, Barger SW. Cytokines and the aging brain – what we don't know might help us. *Trends Neurosci* 2004; **27**: 621–26.
- 119 Weindruch R, Kayo T, Lee CK, Prolla TA. Gene-expression profiling of aging using DNA microarrays. *Mech Ageing Dev* 2002; **123**: 177–93.
- 120 Galvin JE, Ginsberg SD. Expression profiling in the aging brain: a perspective. *Ageing Res Rev* 2005; **4**: 529–47.
- 121 Terao A, Apte-Deshpande A, Dousman L *et al*. Immune response gene expression increases in the aging murine hippocampus. *J Neuroimmunol* 2002; **132**: 99–112.
- 122 Lynch AM, Lynch MA. The age-related increase in IL-1 type I receptor in rat hippocampus is coupled with an increase in caspase-3 activation. *Eur J Neurosci* 2002; **15**: 1779–88.
- 123 Lynch MA. What is the biological significance of an age-related increase in IL-1beta in hippocampus? *Mol Psychiatry* 1999; **4**: 15–18.
- 124 Murray CA, Lynch MA. Evidence that increased hippocampal expression of the cytokine interleukin-1 beta is a common trigger for age- and stress-induced impairments in long-term potentiation. *J Neurosci* 1998; **18**: 2974–81.
- 125 Nimmerjahn A, Kirchhoff F, Helmchen F. Resting microglial cells are highly dynamic surveillants of brain parenchyma in vivo. *Science* 2005; **308**: 1314–18.
- 126 Raivich G. Like cops on the beat: the active role of resting microglia. *Trends Neurosci* 2005; **28**: 571–73.
- 127 Hanisch UK, Kettenmann H. Microglia: active sensor and versatile effector cells in the normal and pathologic brain. *Nat Neurosci* 2007; **10**: 1387–94.
- 128 Ferrari D, Villalba M, Chiozzi P, Ricciardi-Castagnoli P, Di Virgilio F. Mouse microglial cells express a plasma membrane pore gated by extracellular ATP. *J Immunol* 1996; **156**: 1531–39.
- 129 Boddeke EW, Meigel I, Frentzel S, Biber K, Renn LQ, Gebicke-Härter P. Functional expression of the fractalkine (CX3C) receptor and its regulation by lipopolysaccharide in rat microglia. *Eur J Pharmacol* 1999; **374**: 309–13.
- 130 Hanisch UK. Microglia as a source and target of cytokines. *Glia* 2002; **40**: 140–55.
- 131 Davalos D, Grutzendler J, Yang G *et al*. ATP mediates rapid microglial response to local brain injury in vivo. *Nat Neurosci* 2005; **8**: 752–58.
- 132 Becher B, Prat A, Antel JP. Brain-immune connection: immuno-regulatory properties of CNS-resident cells. *Glia* 2000; **29**: 293–304.
- 133 Ekdahl CT, Claassen JH, Bonde S, Kokaia Z, Lindvall O. Inflammation is detrimental for neurogenesis in adult brain. *Proc Natl Acad Sci USA* 2003; **100**: 13632–637.
- 134 Monje ML, Toda H, Palmer TD. Inflammatory blockade restores adult hippocampal neurogenesis. *Science* 2003; **302**: 1760–65.
- 135 Gemma C, Bachstetter AD, Cole MJ, Fister M, Hudson C, Bickford PC. Blockade of caspase-1 increases neurogenesis in the aged hippocampus. *Eur J Neurosci* 2007; **26**: 2795–803.
- 136 Griffin R, Nally R, Nolan Y, McCartney Y, Linden J, Lynch MA. The age-related attenuation in long-term potentiation is associated with microglial activation. *J Neurochem* 2006; **99**: 1263–72.
- 137 Casolini P, Catalani A, Zuena AR, Angelucci L. Inhibition of COX-2 reduces the age-dependent increase of hippocampal inflammatory markers, corticosterone secretion, and behavioral impairments in the rat. *J Neurosci Res* 2002; **68**: 337–43.
- 138 Wilson CJ, Finch CE, Cohen HJ. Cytokines and cognition—the case for a head-to-toe inflammatory paradigm. *J Am Geriatr Soc* 2002; **50**: 2041–56.
- 139 Lynch MA. Analysis of the mechanisms underlying the age-related impairment in long-term potentiation in the rat. *Rev Neurosci* 1998; **9**: 169–201.
- 140 Maher FO, Martin DS, Lynch MA. Increased IL-1beta in cortex of aged rats is accompanied by downregulation of ERK and PI-3 kinase. *Neurobiol Aging* 2004; **25**: 795–806.
- 141 Nolan Y, Maher FO, Martin DS *et al*. Role of interleukin-4 in regulation of age-related inflammatory changes in the hippocampus. *J Biol Chem* 2005; **280**: 9354–62.
- 142 Perry VH, Matyszak MK, Fearn S. Altered antigen expression of microglia in the aged rodent CNS. *Glia* 1993; **7**: 60–67.
- 143 Kullberg S, Aldskogius H, Ulfhake B. Microglial activation, emergence of ED1-expressing cells and clusterin upregulation in the aging rat CNS, with special reference to the spinal cord. *Brain Res* 2001; **899**: 169–86.
- 144 Stichel CC, Luebbert H. Inflammatory processes in the aging mouse brain: participation of dendritic cells and T-cells. *Neurobiol Aging* 2007; **28**: 1507–21.
- 145 Schindler MK, Forbes ME, Robbins ME *et al*. Aging-dependent changes in the radiation response of the adult rat brain. *Int J Radiat Oncol Biol Phys* 2008; **70**: 826–34.

- 146 Sloane JA, Hollander W, Moss MB, Rosene DL, Abraham CR. Increased microglial activation and protein nitration in white matter of the aging monkey. *Neurobiol Aging* 1999; **20**: 395–405.
- 147 Conde JR, Streit WJ. Microglia in the aging brain. *J Neuropathol Exp Neurol* 2006; **65**: 199–203.
- 148 Simpson JE, Ince PG, Higham CE *et al*. Microglial activation in white-matter lesions and nonlesional white matter of ageing brains. *Neuropathol Appl Neurobiol* 2007; **33**: 670–83.
- 149 Long JM, Kalebica AN, Muth NJ *et al*. Stereological analysis of astrocyte and microglia in aging mouse hippocampus. *Neurobiol Aging* 1998; **19**: 497–503.
- 150 Streit WJ. Microglia and neuroprotection: implications for Alzheimer's disease. *Brain Res Brain Res Rev* 2005; **48**: 234–39.
- 151 Ziv Y, Ron N, Butovsky O *et al*. Immune cells contribute to the maintenance of neurogenesis and spatial learning abilities in adulthood. *Nat Neurosci* 2006; **9**: 268–75.
- 152 Glezer I, Simard AR, Rivest S. Neuroprotective role of the innate immune system by microglia. *Neuroscience* 2007; **147**: 867–83.
- 153 Nakajima K, Tohyama Y, Maeda S, Kohsaka S, Kurihara T. Neuronal regulation by which microglia enhance the production of neurotrophic factors for GABAergic, catecholaminergic, and cholinergic neurons. *Neurochem Int* 2007; **50**: 807–820.
- 154 Lai AY, Todd KG. Differential regulation of trophic and proinflammatory microglial effectors is dependent on severity of neuronal injury. *Glia* 2008; **56**: 259–70.
- 155 Sierra A, Gottfried-Blackmore AC, McEwen BS, Bulloch K. Microglia derived from aging mice exhibit an altered inflammatory profile. *Glia* 2007; **55**: 412–24.
- 156 Joseph JA, Shukitt-Hale B, Casadesus G, Fisher D. Oxidative stress and inflammation in brain aging: nutritional considerations. *Neurochem Res* 2005; **30**: 927–35.
- 157 Roth GS, Ingram DK, Joseph JA. Nutritional interventions in aging and age-associated diseases. *Ann N Y Acad Sci* 2007; **1114**: 369–71.
- 158 Bickford PC, Gould T, Briederick L *et al*. Antioxidant-rich diets improve cerebellar physiology and motor learning in aged rats. *Brain Res* 2000; **866**: 211–17.
- 159 Cartford MC, Gemma C, Bickford PC. Eighteen-month-old Fischer 344 rats fed a spinach-enriched diet show improved delay classical eyeblink conditioning and reduced expression of tumor necrosis factor alpha (TNFalpha) and TNFbeta in the cerebellum. *J Neurosci* 2002; **22**: 5813–16.
- 160 Goyarzu P, Malin DH, Lau FC *et al*. Blueberry-supplemented diet: effects on object recognition memory and nuclear factor-kappa B levels in aged rats. *Nutr Neurosci* 2004 Apr; **7**: 75–83.
- 161 Shukitt-Hale B, Carey A, Simon L, Mark DA, Joseph JA. Effects of Concord grape juice on cognitive and motor deficits in aging. *Nutrition* 2006; **22**: 295–302.
- 162 Kamel NS, Gammack J, Cepeda O, Flaherty JH. Antioxidants and hormones as anti-aging therapies: high hopes, disappointing results. *Cleve Clin J Med* 2006; **73**: 1049–56.
- 163 Grodstein F, Kang JH, Glynn RJ, Cook NR, Gaziano JM. A randomized trial of beta carotene supplementation and cognitive function in men: the Physicians' Health Study II. *Arch Intern Med* 2007; **167**: 2184–80.
- 164 McNeill G, Avenell A, Campbell MK *et al*. Effect of multivitamin and multiminerall supplementation on cognitive function in men and women aged 65 years and over: a randomised controlled trial. *Nutr J* 2007; **6**: 10.
- 165 Rondanelli M, Trotti R, Opizzi A, Solerte SB. Relationship among nutritional status, pro/antioxidant balance and cognitive performance in a group of free-living healthy elderly. *Minerva Med* 2007; **98**: 639–45.
- 166 Wengreen HJ, Munger RG, Corcoran CD *et al*. Antioxidant intake and cognitive function of elderly men and women: the Cache County Study. *J Nutr Health Aging* 2007; **11**: 230–37.
- 167 Duffy KB, Spangler EL, Devan BD *et al*. A blueberry-enriched diet provides cellular protection against oxidative stress and reduces a kainate-induced learning impairment in rats. *Neurobiol Aging*. 2007 May 22; [Epub ahead of print]
- 168 Shukitt-Hale B, Carey AN, Jenkins D, Rabin BM, Joseph JA. Beneficial effects of fruit extracts on neuronal function and behavior in a rodent model of accelerated aging. *Neurobiol Aging* 2007; **28**: 1187–94.
- 169 Bekker AY, Weeks EJ. Cognitive function after anaesthesia in the elderly. *Best Pract Res Clin Anaesthesiol* 2003; **17**: 259–72.
- 170 Cohendy R, Brougere A, Cuivillon P. Anaesthesia in the older patient. *Curr Opin Clin Nutr Metab Care* 2005; **8**: 17–1.
- 171 Levine WC, Mehta V, Landesberg G. Anesthesia for the elderly: selected topics. *Curr Opin Anaesthesiol* 2006; **19**: 320–24.
- 172 Culley DJ, Xie Z, Crosby G. General anesthetic-induced neurotoxicity: an emerging problem for the young and old? *Curr Opin Anaesthesiol* 2007; **20**: 408–13.
- 173 Perouansky M. General anesthetics and long-term neurotoxicity. *Handb Exp Pharmacol* 2008; **182**: 143–57.

- 174 Kyrkanides S, O'Banion MK, Whiteley PE, Daeschner JC, Olschowka JA. Enhanced glial activation and expression of specific CNS inflammation-related molecules in aged versus young rats following cortical stab injury. *J Neuroimmunol* 2001; **119**: 269–77.
- 175 Popa-Wagner A, Badan I, Walker L, Groppa S, Patrana N, Kessler C. Accelerated infarct development, cytogenesis and apoptosis following transient cerebral ischemia in aged rats. *Acta Neuropathol (Berl)* 2007; **113**: 277–93.
- 176 Lamproglou I, Chen QM, Boisserie G *et al*. Radiation-induced cognitive dysfunction: an experimental model in the old rat. *Int J Radiat Oncol Biol Phys* 1995; **31**: 65–70.
- 177 Lamproglou I, Baillet F, Boisserie G, Mazon JJ, Delattre JY. The influence of age on radiation-induced cognitive deficit: experimental studies on brain irradiation of 30 Gy in 10 sessions and 12 hours in the Wistar rat at 1 1/2, 4 and 18 months age. *Can J Physiol Pharmacol* 2002; **80**: 679–85.
- 178 Abrey LE, Deangelis LM, Yahalom J. Long-term survival in primary CNS lymphoma. *J Clin Oncol* 1998; **16**: 859–63.
- 179 Deangelis LM. Primary CNS lymphoma: treatment with combined chemotherapy and radiotherapy. *J Neurooncol* 1999; **43**: 249–57.
- 180 Swennen MH, Bromberg JE, Witkamp TD, Terhaard CH, Postma TJ, Taphoorn MJ. Delayed radiation toxicity after focal or whole brain radiotherapy for low-grade glioma. *J Neurooncol* 2004; **66**: 333–39.
- 181 Omuro AM, Ben-Porat LS, Panageas KS, *et al*. Delayed neurotoxicity in primary central nervous system lymphoma. *Arch Neurol* 2005; **62**: 1595–1600.
- 182 Barrientos RM, Higgins EA, Biedenkapp JC *et al*. Peripheral infection and aging interact to impair hippocampal memory consolidation. *Neurobiol Aging* 2006; **27**: 723–32.
- 183 Frank MG, Baratta MV, Sprunger DB, Watkins LR, Maier SF. Microglia serve as a neuroimmune substrate for stress-induced potentiation of CNS pro-inflammatory cytokine responses. *Brain Behav Immun* 2007; **21**: 47–59.
- 184 Perry VH, Cunningham C, Holmes C. Systemic infections and inflammation affect chronic neurodegeneration. *Nat Rev Immunol* 2007; **7**: –67.
- 185 Chen J, Buchanan JB, Sparkman NL, Godbout JP, Freund GG, Johnson RW. Neuroinflammation and disruption in working memory in aged mice after acute stimulation of the peripheral innate immune system. *Brain Behav Immun* 2008; **3**: 301–11.
- 186 Mangano EN, Hayley S. Inflammatory priming of the substantia nigra influences the impact of later paraquat exposure: neuroimmune sensitization of neurodegeneration. *Neurobiol Aging* 2008 (in press).
- 187 Viviani B, Gardoni F, Marinovich M. Cytokines and neuronal ion channels in health and disease. *Int Rev Neurobiol* 2007; **82**: 247–63.
- 188 Pickering M, Cumiskey D, O'Connor JJ. Actions of TNF-alpha on glutamatergic synaptic transmission in the central nervous system. *Exp Physiol* 2005; **90**: 663–70.
- 189 Pickering M, O'Connor JJ. Pro-inflammatory cytokines and their effects in the dentate gyrus. *Prog Brain Res* 2007; **163**: 339–54.
- 190 Neumann H, Schweigreiter R, Yamashita T, Rosenkranz K, Wekerle H, Barde YA. Tumor necrosis factor inhibits neurite outgrowth and branching of hippocampal neurons by a rho-dependent mechanism. *J Neurosci* 2002; **22**: 854–62.
- 191 Katafuchi T, Take S, Hori T. Roles of cytokines in the neural-immune interactions: modulation of NMDA responses by IFN-alpha. *Neurobiology* 1995; **3**: 319–27.
- 192 Vereyken EJ, Bajova H, Chow S, de Graan PN, Gruol DL. Chronic interleukin-6 alters the level of synaptic proteins in hippocampus in culture and in vivo. *Eur J Neurosci* 2007; **25**: 3605–16.
- 193 Bessis A, Béchade C, Bernard D *et al*. Microglial control of neuronal death and synaptic properties. *Glia* 2007; **55**: 233–38.
- 194 Trapp BD, Wujek JR, Criste GA *et al*. Evidence for synaptic stripping by cortical microglia. *Glia* 2007; **55**: 360–68.

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